

Transcutaneous CO₂ tension measurement as an indicator of severity of hemorrhagic shock

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Abstract: This study was undertaken to evaluate whether transcutaneous CO₂ tension (PtcCO₂) could be used as an indicator of the global systemic severity of hemorrhagic shock. PtcCO₂ levels in ten anesthetized mongrel dogs were measured during hemorrhage and during volume restoration and were correlated with mixed venous CO₂ tension (P \bar{v} CO₂). After withdrawal of 30 ml·kg⁻¹ blood, both PtcCO₂ and P \bar{v} CO₂ increased significantly (from 43 ± 7 to 70 ± 27 torr ($P < 0.05$) and from 48 ± 6 to 59 ± 12 torr ($P < 0.05$), respectively). Throughout the experiments, PtcCO₂ levels changed almost in parallel to P \bar{v} CO₂ levels. However, changes in PtcCO₂ exceeded those in P \bar{v} CO₂ from the end of hemorrhage, at which time cardiac output decreased to 35% of the baseline value, until the end of volume restoration, and the changes in PtcCO₂ showed a close logarithmic relationship with P \bar{v} CO₂ ($r = 0.78$, $n = 110$). Additionally, arterio-transcutaneous CO₂ tension gradients (P[tc- \bar{v}]CO₂) showed a close exponential correlation with cardiac output per body weight (CO/BW) during the shedding phase ($r = 0.85$, $n = 60$), although the correlation with CO/BW lessened during the retransfusion phase ($r = 0.55$, $n = 60$). PtcCO₂ was roughly correlated with P \bar{v} CO₂ during hemorrhagic shock, and levels of PtcCO₂ higher than P \bar{v} CO₂ reflected critical tissue perfusion.

Key words: Transcutaneous CO₂ tension, Mixed venous CO₂ tension, Hemorrhagic shock

Introduction

Venous and tissue hypercarbia is provoked during circulatory and other forms of shock, and has been related to reduced CO₂ clearance due to decreased pulmonary vascular flow, as well as to increased CO₂ generation due to the buffering of increased hydrogen ion production in the hypoxic cell [1]. It is now well recognized that

mixed venous CO₂ tension (P \bar{v} CO₂) values indicate changes in tissue perfusion [2–5]. In recent years, several studies [6–9] have focused on increases in gastrointestinal mucosal CO₂ tension, which can be measured utilizing a tonometer or an ion-sensitive field-effect transistor (ISFET) sensor, during low-flow conditions. These studies have reported that PCO₂ values are reliable indicators of tissue hypoxia and a clinically useful predictor of prognosis. However, it is often difficult to obtain mixed venous blood from patients in shock or from pediatric patients, and the tonometric system cannot provide continuous information. Furthermore, the ISFET sensor system, in which a micro-sensor is surgically inserted into the submucosal tissue, is invasive and has not yet been clinically established. A noninvasive and continuous method of monitoring tissue CO₂ levels is needed to manage unstable patients during shock.

Transcutaneous CO₂ tension (PtcCO₂) monitoring has been widely accepted, particularly in neonatal intensive care, since it was developed by Huch et al. [10] in 1973. Animal experimental studies have shown that PtcCO₂ correlates directly with arterial CO₂ tension (PaCO₂) in normal circulation and correlates inversely with the cardiac index during low-flow shock [11]. These studies utilized very severe shock brought about by sudden withdrawal of blood, but the characteristics in moderate shock models have not yet been fully evaluated.

In the present study, we utilized gradient withdrawal of the blood to create the model and reconsidered whether PtcCO₂ could be an indicator of the global systemic severity of shock.

Materials and methods

Experimental preparation

The study protocol was reviewed and approved by the Institutional Animal Care and Use Committee. Ten

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adult mongrel dogs, weighing 7.6–13.4 kg (mean, 9.7 kg) were used. Anesthesia was induced with intramuscular ketamine (10 mg·kg⁻¹) and succinylcholine (2 mg·kg⁻¹), and was maintained with 1% isoflurane. After endotracheal intubation, ventilation was initiated with a Harvard animal respirator (Model B2; Igarashi, Tokyo, Japan), using 30% oxygen and a tidal volume of 10 ml·kg⁻¹. The respiration rate was adjusted to 10–20 min⁻¹ so that the end-tidal CO₂ tension was maintained at 35 torr, measured with an expired CO₂ monitor (Respina IH31; San-ei, Tokyo, Japan). The settings of the respirator were not changed thereafter.

Experimental protocol

An endovascular catheter was inserted into the jugular vein and the right carotid artery for blood shedding and blood sampling. A 5-Fr thermodilution catheter was also inserted via the right jugular vein. After systemic heparinization (5 mg·kg⁻¹ intravenously), blood was withdrawn at a rate of 6 ml·kg⁻¹·5 min⁻¹. Blood was withdrawn five times, at 5-min intervals, and a total of 30 ml·kg⁻¹ was removed. Subsequently, the blood was restored at the same rate.

Measurements

Four parameters were measured after each cycle of hemorrhagic shock and volume restoration: transcutaneous gas tension, arterial gas tension, mixed venous gas tension, and cardiac output. Transcutaneous gas tension was measured with a transcutaneous gas monitor (Models 632 and 634; Kontron, Everett, MA, USA). The probes were warmed to 43°C. The oxygen probe was calibrated with air, and the CO₂ probe was calibrated with 5% and 10% CO₂. The probes were then placed on the shaved anterior chest wall. Gas tension in arterial blood and mixed venous blood was measured with an automatic blood gas analyzer (Stat Profile 5; Novamatrix Medical System, Wallingford, CT, USA) immediately after blood sampling. Cardiac output was

measured by the thermodilution method, using a cardiac output computer (9520A; Baxter Edward Laboratory, Irvine, CA, USA).

Statistical analysis

The measured variables were evaluated by analysis of variance (two-factor) with repeated measures, followed by Bonferroni's correction for multiple comparisons. Regression analysis and calculation of correlation coefficients were performed for PtcCO₂, PaCO₂, and P \bar{v} CO₂, as well as for cardiac output per body weight (CO/BW) and arterio-transcutaneous CO₂ tension gradients (P[tc-a]CO₂), at each point of measurement. Differences were considered significant at $P < 0.05$.

Results

Changes in CO/BW

CO/BW decreased to 35% of the baseline value by gradient hemorrhage of 30 ml·kg⁻¹·50 min⁻¹ and recovered to the baseline value on the retransfusion of 24 ml·kg⁻¹·40 min⁻¹. After the retransfusion of all shed blood, CO/BW increased to 112% of the baseline value (Table 1).

Changes in PaCO₂, P \bar{v} CO₂, and PtcCO₂ (Table 1, Fig. 1)

PaCO₂ remained relatively unchanged during hemorrhage and then rose slightly after volume restoration.

The mean baseline P \bar{v} CO₂, of 48 ± 6 torr, began rising when CO/BW was approximately 50% of its baseline value. P \bar{v} CO₂ reached a peak 20 min after the beginning of volume restoration, increasing to 59 ± 12 torr ($P < 0.05$) and then decreasing to the pre-shock level.

PtcCO₂ changed almost in parallel to P \bar{v} CO₂. The mean baseline PtcCO₂ was 43 ± 7 torr, and the value increased to 70 ± 27 torr ($P < 0.05$) 10 min after the

Table 1. Values of all variables measured during hemorrhage and retransfusion (means ± SD)

	Baseline		Hemorrhage					Retransfusion				
	0 min	10 min	20 min	30 min	40 min	50 min	60 min	70 min	80 min	90 min	100 min	
CO/BW (ml·min ⁻¹ ·kg ⁻¹)	158 ± 31	135 ± 26	112 ± 25**	92 ± 25**	74 ± 18**	56 ± 21**	84 ± 28**	112 ± 36**	136 ± 34	159 ± 39	177 ± 46	
PaO ₂ (torr)	118 ± 17	115 ± 19	114 ± 20	112 ± 21	115 ± 19	117 ± 18	105 ± 22	102 ± 24	103 ± 27	103 ± 23	100 ± 20	
P \bar{v} O ₂ (torr)	49 ± 6	45 ± 7	41 ± 6*	37 ± 8**	33 ± 6**	28 ± 8**	34 ± 7**	41 ± 8*	45 ± 8	47 ± 8	50 ± 7	
PtcO ₂ (torr)	97 ± 16	91 ± 15	84 ± 16	72 ± 17**	53 ± 21**	36 ± 20**	46 ± 24**	58 ± 22**	70 ± 21**	79 ± 19	84 ± 17	
PaCO ₂ (torr)	39 ± 5	38 ± 6	38 ± 7	38 ± 7	38 ± 8	36 ± 8	42 ± 11	45 ± 11	44 ± 10	44 ± 10	43 ± 10	
P \bar{v} CO ₂ (torr)	48 ± 6	47 ± 5	47 ± 6	48 ± 9	51 ± 7	53 ± 7	58 ± 11*	59 ± 12*	55 ± 10	51 ± 11	50 ± 9	
PtcCO ₂ (torr)	43 ± 7	42 ± 8	43 ± 9	46 ± 12	51 ± 16	61 ± 23	70 ± 27*	69 ± 29*	64 ± 28	59 ± 23	54 ± 19	
P(\bar{v} -a)CO ₂ (torr)	8 ± 4	9 ± 3	9 ± 4	10 ± 5	14 ± 6	17 ± 8*	17 ± 7*	14 ± 8	11 ± 5	7 ± 6	7 ± 5	
P(tc-a)CO ₂ (torr)	3 ± 4	4 ± 4	5 ± 6	8 ± 8	14 ± 12*	25 ± 19**	28 ± 21**	25 ± 23*	20 ± 21	14 ± 17	11 ± 15	

* $P < 0.05$; ** $P < 0.01$ compared with baseline values.

CO/BW, cardiac output per body weight; P(\bar{v} -a)CO₂, arteriovenous PCO₂ gradient; P(tc-a)CO₂, arterio-transcutaneous PCO₂ gradient.

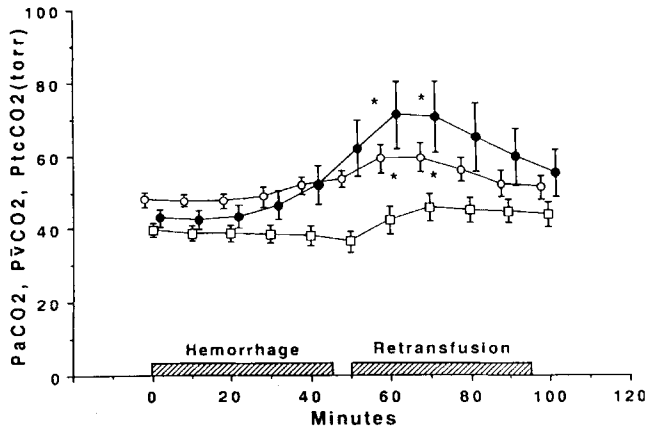
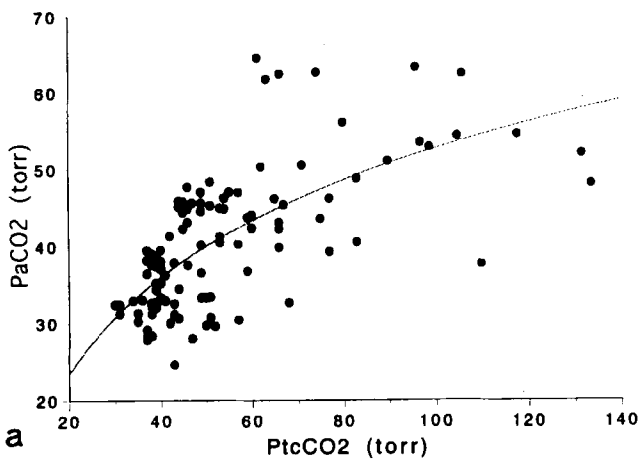


Fig. 1. Changes in PaCO₂ (squares), mixed venous CO₂ tension (PvCO₂; circles) and transcutaneous CO₂ tension (PtcCO₂; dots) during hemorrhagic and retransfusion. **P* < 0.05 compared with baseline values. Values are means ± SEM

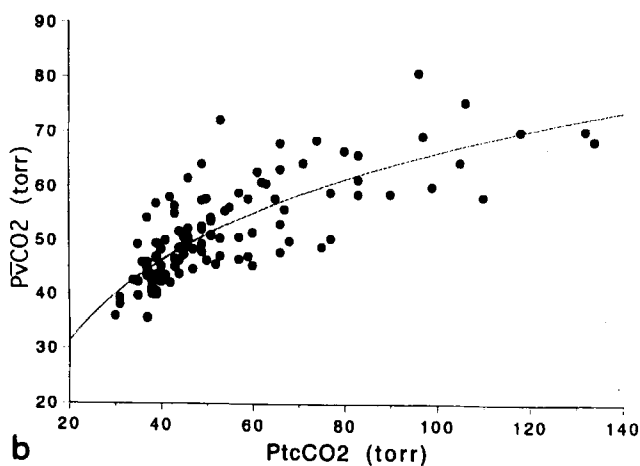
beginning of volume restoration. Thus, in the early phase of hemorrhage, the PtcCO₂ value lay between the values for PaCO₂ and PvCO₂, but it subsequently increased, exceeding PvCO₂ and remaining higher until the end of volume restoration. PtcCO₂ did not return to control values until 20min after the recovery of cardiac output.

Relationship of PaCO₂, PvCO₂, and PtcCO₂

Throughout this experiment, PtcCO₂ showed a closer logarithmic correlation to PvCO₂ (*r* = 0.78, *n* = 110) than to PaCO₂ (*r* = 0.68, *n* = 110) (Fig. 2). Additionally, in several cases, PtcCO₂ values increased to about two-fold over PvCO₂ values from the end of hemorrhagic shock until the early phase of volume restoration, although there was no significant difference between PtcCO₂ and PvCO₂ (Table 1, Fig. 2).

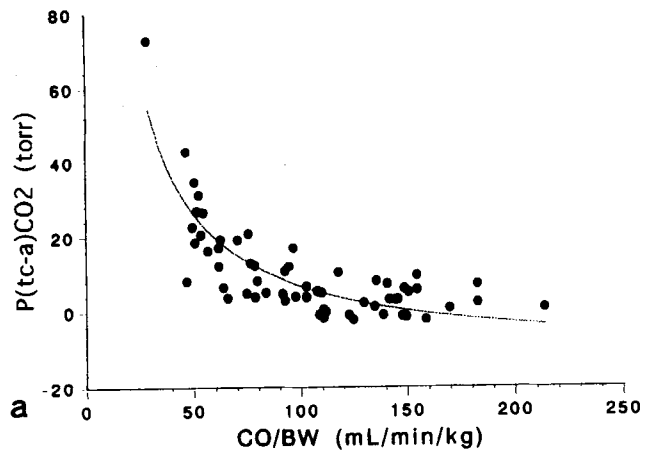


a

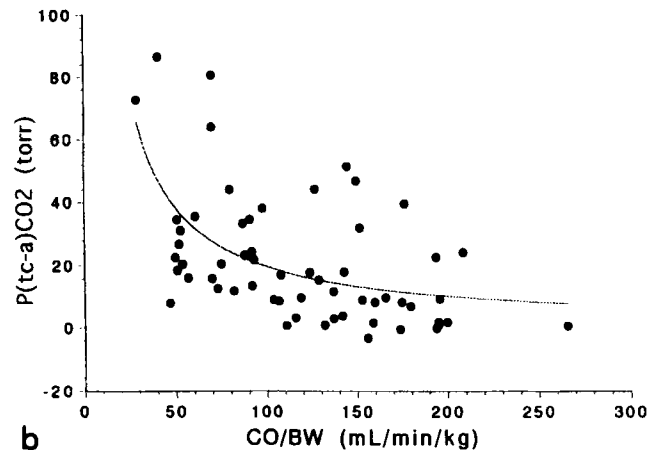


b

Fig. 2a,b. Relationship between PtcCO₂ and both **a** PaCO₂ and **b** PvCO₂ during hemorrhage and retransfusion. **a**, $y = 18 * \ln(x) - 30.5$; *n* = 110; *r* = 0.68; **b**, $y = 21.4 * \ln(x) - 32.9$; *n* = 110; *r* = 0.78



a



b

Fig. 3a,b. Relationship between cardiac output per body weight (CO/BW) and arterio-transcutaneous PCO₂ gradient (P[tc-a]CO₂) during **a** hemorrhage and **b** retransfusion. **a**, $y = 1401/x - 12.18$; *n* = 60; *r* = 0.85; **b**, $y = 1872/x + 0.8$; *n* = 60; *r* = 0.55

Relationship of P(tc-a)CO₂ and CO/BW

P(tc-a)CO₂ showed a close exponential correlation with CO/BW during the shedding phase ($r = 0.85$, $n = 60$). However, the correlation with CO/BW lessened during the retransfusion phase ($r = 0.55$, $n = 60$) (Fig. 3).

Discussion

A major finding of the present experimental study was the close logarithmic correlation between PtcCO₂ and P \bar{v} CO₂. This suggests that PtcCO₂, as well as P \bar{v} CO₂, reflects whole body hypercarbia fairly well during hemorrhagic shock. However, as cardiac output was reduced, values for PtcCO₂ differed from both PaCO₂ and P \bar{v} CO₂ values, remaining in excess of P \bar{v} CO₂ until the end of volume restoration. These changes may reflect two factors, the first being the disturbed distribution of blood flow to the various organs caused by the progression of shock. Previous studies indicate that, of all the vascular beds, the cutaneous vascular bed exhibits the greatest vasoconstrictive response and the most severe blood flow reduction during progressive blood loss, since this vascular bed possesses no local intrinsic blood flow regulation (i.e., autoregulation) and is under strong extrinsic (e.g., neural/humoral) control [12]. During circulatory shock, the skin is one of the first organs in which blood flow reduction occurs and the skin is also among the last organs to be restored to metabolic balance during resuscitation from shock [13–15]. Accordingly, the measurement of PtcCO₂ may be advantageous for the early diagnosis and assessment of shock, although this method may overestimate the extent of systemic injury.

The second factor that could explain the discrepancy between PtcCO₂ and P \bar{v} CO₂ values is a reduction in the washout of tissue gases into venous blood due to severe peripheral circulatory impairment. From this perspective, not only the conventional analysis of arterial gases but also the analysis of venous gases has limited value in the presence of severe shock and does not necessarily provide direct information about the global adequacy of oxygenation. In other words, only the monitoring of tissue gases can warn of critical conditions.

Johnson and Weil [1] found an exponential correlation between arteriovenous PCO₂ gradient (P[\bar{v} -a]CO₂) and cardiac output during hypoperfusion, using the Fick equation. This was in accord with the correlation between P(tc-a)CO₂ and CO/BW found in the present study. P(tc-a)CO₂ should provide an indication of the severity of inadequate cutaneous perfusion analogous to the arteriovenous PCO₂ gradient during global hypoperfusion [3–5]. As shown in Fig. 3, the curve increased steeply at approximately 50–60 ml·min⁻¹·kg⁻¹. According to the method described by Samsel and

Schumacker [16], this point may be a critical value for skin, and seems to correspond to the point at which PtcCO₂ became higher than P \bar{v} CO₂ during the shedding phase.

Several other studies have focused on increases in PCO₂ in various tissues, such as the gastrointestinal (GI) tract [6–9], the myocardium [17], the brain [18], and the muscle [19] as an indicator of the adequacy of tissue oxygenation. Hypercarbia in various tissues appears to be the predominant phenomenon during hypoperfusion. PCO₂ in GI mucosa has been particularly well investigated during various types of shock, such as hemorrhagic [8,9], septic [20], cardiogenic [7], and anaphylactic [9], and this parameter has been reported to be an extremely sensitive indicator of systemic damage, since the GI mucosa is quite vulnerable to even mild hypoxia.

Tang and coworkers (Noc et al. [8] and Tang et al. [9]) monitored gastric intramural PCO₂ during hemorrhage and volume restoration, utilizing an ISFET sensor. The changes they reported in gastric intramural PCO₂ were in approximate agreement with those we found in PtcCO₂ in the present experiment. It appears that PtcCO₂ monitoring compares favorably with GI mucosal PCO₂ monitoring in terms of precision and reliability, at least during hemorrhage and volume restoration. Moreover, PtcCO₂ monitoring is a noninvasive and clinically well established method, this constituting an advantage over the ISFET sensor system. Further study is required to determine whether the present observations apply to other types of shock.

The monitoring of PtcCO₂ entails several problems for accurate interpretation of hypercarbia throughout the body. One problem arises from the disturbed distribution of blood flow among various tissues, as described above. This disturbed distribution is also seen among various skin regions [21]. An additional problem is related to measurement techniques. When the probe is heated, measurements can be altered by arterIALIZATION of the capillaries, by CO₂ production following metabolic enhancement, by changes in the CO₂ transporting capacity, and by diffusion resistance [10,22,23]. Hence, it should be considered that the values obtained while hypoperfusion is monitored may vary widely.

In conclusion, PtcCO₂ roughly correlated with P \bar{v} CO₂ during hemorrhagic shock, and PtcCO₂ levels higher than P \bar{v} CO₂ levels reflected critical tissue perfusion. P \bar{v} CO₂ monitoring provided more precise information about global systemic change than P \bar{v} CO₂ analysis, particularly in conditions of severe low blood flow.

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References

1. Johnson BA, Weil MH (1991) Redefining ischemia due to circulatory failure as dual defects of oxygen deficits and of carbon dioxide excesses. *Crit Care Med* 19:1432–1438
2. Grundler W, Weil MH, Rackow EC (1986) Arteriovenous carbon dioxide and pH gradients during cardiac arrest. *Circulation* 74:1071–1074
3. Ducey JP, Lamiell JM, Gueller GE (1992) Arterial-venous carbon dioxide tension difference during severe hemorrhage and resuscitation. *Crit Care Med* 20:518–522
4. Zhang H, Vincent J (1993) Arteriovenous differences in PCO₂ and pH: Good indicators of critical hypoperfusion. *Am Rev Respir Dis* 148:867–871
5. Linden PV, Rausin I, Deltell A, Bekrar Y, Gilbert E, Bakker J, Vincent JL (1995) Detection of tissue hypoxia by arteriovenous gradient for PCO₂ and pH in anesthetized dogs during progressive hemorrhage. *Anesth Analg* 80:269–275
6. Fiddian-Green RG, Pittenger BS, Whitehouse WM (1982) Back-diffusion of CO₂ and its influence on the intramural pH in gastric mucosa. *J Surg Res* 33:39–48
7. Fiddian-Green RG, Amelin PM, Herrmann JB, Arous E, Cutler BS, Schiedler M, Wheeler B, Baker S (1986) Prediction for the development of sigmoid ischemia on the day of aortic operations. *Arch Surg* 121:654–660
8. Noc M, Weil MH, Sun S, Gazmuri RJ, Tang W, Pakula JL (1993) Comparison of gastric luminal and gastric wall PCO₂ during hemorrhagic shock. *Circ Shock* 40:194–199
9. Tang W, Weil MH, Sun S, Noc M, Gazmuri RJ, Bisera J (1994) Gastric intramural PCO₂ as monitor of perfusion failure during hemorrhagic and anaphylactic shock. *J Appl Physiol* 76:572–577
10. Huch A, Lübbers DW, Huch R (1973) Patientenüberwachung durch transcutane PCO₂ messung bei gleichzeitiger kontrolle der relativen lokalen perfusion (in German). *Anaesthesist* 22:379–382
11. Tremper KK, Mentelos RA, Shoemaker WC (1980) Effects of hypercarbia and shock on transcutaneous carbon dioxide at different electrode temperatures. *Crit Care Med* 8:709–713
12. Bond RF (1992) Peripheral macro- and microcirculation. In: Schlag G, Redl H (eds) *Pathophysiology of shock, sepsis, and organ failure*. Springer, Berlin Heidelberg New York Tokyo, pp 893–907
13. Lübbers DW (1981) Theoretical basis of the transcutaneous blood gas measurements. *Crit Care Med* 9:721–733
14. Tremper KK, Shoemaker WC (1981) Transcutaneous oxygen monitoring of critically ill adults, with and without low flow shock. *Crit Care Med* 9:706–709
15. Hartmann M, Montgomery A, Jonsson K, Haglund U (1991) Tissue oxygen in hemorrhagic shock measured as transcutaneous oxygen tension, subcutaneous oxygen tension, and gastrointestinal intramucosal pH in pigs. *Crit Care Med* 19:205–210
16. Samsel RW, Schumacker PT (1988) Determination of the critical O₂ delivery from experimental data: Sensitivity to error. *J Appl Physiol* 64:2074–2082
17. Kette F, Weil MH, Gazmuri RJ, Bisera J, Rackow EC (1993) Intramyocardial hypercarbic acidosis during cardiac arrest and resuscitation. *Crit Care Med* 21:901–906
18. von Hanwehr R, Smith M, Siesjo BK (1986) Extra- and intracellular pH during near-complete forebrain ischemia in the rat. *J Neurochem* 46:331–338
19. Furuse A, Brawley BK, Struve E (1973) Skeletal muscle gas tension: Indicator of cardiac output and peripheral tissue perfusion. *Surgery* 74:214–222
20. Desai VS, Weil MH, Tang W, Yang G, Bisera J (1993) Gastric intramural PCO₂ during peritonitis and shock. *Chest* 104:1254–1258
21. Shime N, Yoshioka M, Fukui M, Hatanaka T, Yan T, Tanaka Y (1994) The usefulness of transcutaneous gas monitoring during hemorrhagic shock: Discrepancy between the two transcutaneous gas tensions of anterior thorax and femur (in Japanese with English abstract). *Masui (Jpn J Anesthesiol)* 43:1174–1178
22. Versmold HT, Brunstler I, Enders A, Graubner U, Kopecky M, Schultze J, Sengespeik C, Wittermann C, Zimmer U (1981) Transcutaneous PCO₂ monitoring of newborn infants in shock at electrode temperatures of 41°C–44°C. *Int Care Med* 7:251–252
23. Eberhard P, Mindt W, Schafer R (1981) Cutaneous blood gas monitoring in the adult. *Crit Care Med* 9:702–705